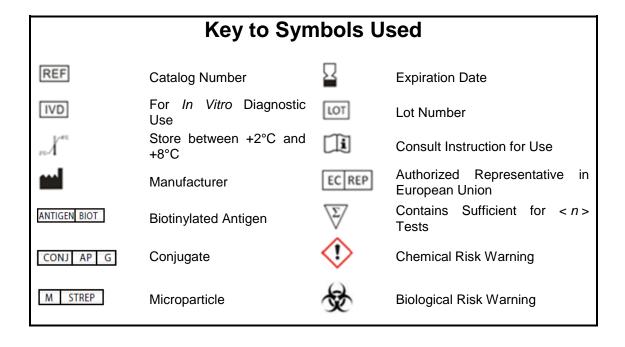
BioCLIA[®] Autoimmune Reagent Kit

DGP IgG

Chemiluminescent Microparticle Immunoassay

Magnetic bead chemiluminescence immunoassay (CLIA) for semi-quantitative determination of anti-DGP IgG antibody in human serum/plasma



BioCLIA® Autoimmune Reagent

Kit, DGP IgG

Intended Use

BioCLIA DGP IgG (Chemiluminescent Microparticle Immunoassay) is intended for the in vitro semi-quantitative measurement of IgG antibodies directed to deamidated gliadin peptide in human serum and plasma as an aid in the diagnosis of celiac disease (CD). This kit is used on the instrument of BioCLIA® 1200 and BioCLIA® 6500.

Catalog Numbers

MY00141 (50 Tests/kit) MY00192 (100 Tests/kit)

Summary and Explanation

Anti-deamidated gliadin peptide antibodies (anti-DGP antibodies) are produced in response to gliadin, a prolamin found in wheat. DGP IgA is found in about 80% of patients with coeliac disease. 1, 2 It is directed against the alpha/beta and gamma (α,β,γ) gliadins. 3 It is also found in a number of patients who are not enteropathic. Some of these patients may have neuropathies that respond favorably to a gluten elimination diet. This is referred to as gluten-sensitive idiopathic neuropathy. ⁴ Clinically these antibodies and IgG antibodies to gliadin are abbreviated as AGA. The IgG antibody is similar to AGA IgA, but is found at higher levels in patients with the IgA-less phenotype. It is also associated with coeliac disease and non-celiac gluten sensitivity. 5-7

AGAs were one of the first serological markers for coeliac disease. Problematic with AGA is the typical sensitivity and specificity was about 85%. Gliadin peptides which are synthesized as the deamidated form have much higher sensitivity and specificity, creating 2 serological tests for CD that approach biopsy diagnostic in performance. ⁸

Principles of the Procedure

BioCLIA DGP IgG assay is a two-step immunoassay using microparticle, enzyme-labeled chemiluminescent technology.

In the first step, the streptavidin coated magnetic microparticle, the biotinylated DGP IgG and human serum/plasma sample are mixed in an assay tube, which allows patient specific anti-DGP IgG to bind. Secondly, after incubation, unbound biotinylated DGP IgG and sample matrix are removed by washing, and the microparticle-DGP IgG-anti-DGP IgG antibodies immune complexes are kept with the help of a magnetic separator. Third, anti-human IgG conjugated alkaline phosphatase is added. Fourth, after incubation, excess enzyme conjugates are removed by washing and finally the bound enzyme is detected by addition of chemiluminescent substrate.

The relative light unit (RLU) intensity is proportional to the amount of anti-DGP IgG antibodies. According to the anti-DGP IgG specific IgG RLU-concentration standard curve, the RLU tested can be interpreted to anti-DGP IgG specific IgG concentration in the sample expressed as RU/mL.

For semi-quantitation of anti-DGP IgG antibodies, the BioCLIA DGP IgG assay utilizes a predefined lot specific Master Curve that is uploaded into the instrument through the reagent Master Calibration Curve barcode. Based on the Master Curve, and results obtained by running two Calibrators, an instrument specific Working Curve is created, which is used to calculate anti-DGP IgG antibodies concentration RU/mL from the relative luminescent units (RLU) obtained for each sample.

Specimen Collection

The appropriate specimen types for BioCLIA DGP IgG is human serum and plasma(Sodium citrate anticoagulant; Heparin anticoagulant; EDTA anticoagulant). Cloudy samples should be purified by low-speed centrifugation. To prevent erroneous results due to the presence of fibrin, ensure that complete clot formation has taken place prior to centrifugation of samples. Some samples, particularly those from patients receiving anticoagulant therapy, may require increasing clotting time.

Freshly collected specimens stored in refrigerator (2-8 °C) are valid for testing within 8 days. The stored specimen should reach to room temperature (18-25°C) before testing, and should not be stored in this temperature condition more than 2 days. All on board specimens should be tested within 10 hours. Three freeze (at -20°C) -thaw cycles for specimens do not affect the testing results.

Warnings and Precautions

- 1. This assay is only for use in the BioCLIA® 1200 and BioCLIA® 6500.
- 2. This product requires the handling of calibrators, controls and human specimens which contain human sourced materials. It is recommended that all human sourced materials are considered to be potentially infectious and handled in accordance with the OSHA Standard on Bloodborne Pathogens. ⁹ Biosafety Level 2 ¹⁰ or other appropriate biosafety practices ^{11, 12}
- 2 or other appropriate biosafety practices should be used for materials that contain or possibly contain infectious agents.
- 3. Liquid waste and solid waste are temporarily stored at separate containers. Waste management should also be handled in accordance with standards mentioned in chapter Warnings and Precautions point No. 2.
- 4. Spilled reagents should be cleaned up immediately. Observe all federal, state and local environmental regulations when disposing wastes.
- 5. Once opened, this reagent cartridge must be stored in the instrument's reagent carousel. Avoid

spilling the reagents when the reagent cartridge is placed into the instrument.

6. Chemical contamination of the reagents can resulting from improper cleaning or rinsing of the instrument. Residues from common laboratory chemicals such as formalin, bleach, ethanol, or detergent can cause interference in the assay. Be sure to follow the recommended cleaning procedure of the instrument as outlined in the BioCLIA® 1200 and BioCLIA® 6500 operator.



7. Precautions:

Proclin 300 added in the reagents (DGP IgG Antigen, Conjugate) at concentration between 0.0015% -0.6%.

Storage Instructions

The kit is stable until the expiration date, if it is stored and handled as directed. Routine store the kit in refrigerator (2-8 °C). Vial opened reagents or onboard reagents can be used up to 56 consecutive days (2-8 °C). The BioCLIA® 1200 and BioCLIA® 6500 software monitors the expiration of the reagent cartridge. The system will not accept expired reagents. Three freeze (at -20°C) -thaw cycles before testing has no effect on the kit reagents.

Materials Supplied

Components are matched in sets. Labels supplied within the kit will be needed for the assay testing. BioCLIA DGP IgG

DGP IgG Antigen 1 bottle (2.5/5 mL) Biotinylated DGP IgG antigen in 0.01 M PBS (pH7.4) DGP-G BIOT buffer with stabilizer.

Preservatives: 0.0015% < Proclin 300 < 0.6%.

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- Conjugate 1 bottle (6.75/13.5 mL) AP labeled anti-human IgG antibodies in 0.05 M MES CONJ AP G (pH6.0) Buffer with stabilizer.
- Microparticle 1 bottle (2.5/5)Streptavidin-microparticles in 0.01 M PBS (pH7.4)

M STREP

Preservatives: 5-Bromo-5-Nitro-1, 3-Dioxane (BND) < 1%.

Kit Component Supplied Separately

buffer with stabilizer.

Additional Materials Required But Not Provided:

- BioCLIA® 1200 (Cat No. MA00139)
- BioCLIA®6500 (Cat No. MA00243)
- BioCLIA Autoimmune Calibrator Set, DGP IgG (Cat No. MY00200, 2 x 1 mL; Cat No. MY00251, 4 x 1 mL)
- BioCLIA Autoimmune Control Set, DGP IgG (Cat No. MY00302, 2 x 1 mL;

- Cat No. MY00353, 4 x 1 mL)
- BioCLIA Sample Diluent I (Cat No. MY00965)
- BioCLIA System Wash Buffer (Cat No. MY00404)
- BioCLIA System Substrate (Cat No. MY00405)
- BioCLIA Cuvettes (Cat No. MA00138, MA00244)
- BioCLIA Silicon Gasket (Small) (Cat No. MV00195)
- BioCLIA Silicon Gasket (Large) (Cat No. MV00196)
- BioCLIA Substrate Tube Maintenance cleanser (Cat No. MA00140)
- BioCLIA Sample Probe Maintenance cleanser (Cat No. MA00141)
- BioCLIA Micro Cup (Cat No. MA00142)
- Distilled Water

Assay Procedure

Note that, it is important to perform all routine maintenance procedures for optimal performance,, such as routine cleaning, calibration and control procedures that are defined in the BioCLIA® 1200 and BioCLIA® 6500 User Manual.

See the BioCLIA® 1200 and BioCLIA® 6500 User Manual for preparation, setup, dilutions, adjustment, assay and quality control procedures.

Users should have the periodic calibration procedure for every 56 consecutive days from last calibration. Besides, a calibration procedure should be carried out when a new batch of BioCLIA DGP IgG kit is used.

The control procedure should be done before running the specimens each day. Users also can adjust the control procedure period according to their own lab frequency.

Expected Values

Each Laboratory should establish its own reference

When the customer see a problem (High CV or unusual values, rerun controls and analyze specimens again.

Result Analysis

With the help of the built in master calibration curve and specified two-point calibrator set for the instrument, the BioCLIA will automatically calculate the auto-antibodies concentration of each specimen and interpret the results into RU/mL. The concentration of anti-DGP IgG antibody sample is reported as < 2 RU/mL when it is lower than the minimum detection limit, while reported as > 400 RU/mL when it is higher than the range of measurement.

Sample Dilution

The specimens are diluted with BioCLIA Sample Diluent I before testing (dilution ratio 1:20) by the BioCLIA® 1200 and BioCLIA® 6500 automatically.

Cut-Off Value Determination

120 clinical samples, including 30 positive sera, 30

negative sera, 30 positive plasma and 30 negative plasma were collected and valued. These samples were venous blood from human aged between 0 - 80, sealed and stored at 2 - 8 °C. Results of 120 clinical samples tested by the BioCLIA DGP IgG kit were analyzed using the receiver-operating characteristic curve (ROC) and the cut-off value was determined at 20 RU/mL.

Test Result Interpretation

Specimen with concentration < 20 RU/mL, interpreted as negative;

Specimen with concentration ≥ 20 RU/mL, interpreted as positive.

Test results only reflect the sample collecting status and should be interpreted/analyzed for diagnosis in conjunction with other laboratory and clinical findings.

Performance Characteristics

APPEARENCE

Kit components are complete with no leakage. No precipitation or floc in liquid reagents. Packing labels are clear and easy to be identified.

ACCURACY / SPIKED RECOVERY

The accuracy/spiked recovery was determined by analyzing samples spiked with known amounts of anti-DGP IgG antibodies into certain matrix. Anti-DGP IgG antibody positive samples (low 100 RU/mL, mid 200 RU/mL, high 300 RU/mL) were spiked into two matrixes (50 & 100 RU/mL) separately at the volume ratio of 1:9, making totally 6 spiked samples and each sample was tested in triplicate. The spiked recovery for the concentration of anti-DGP IgG antibodies was calculatd.*

Calculatu.						
	Matrix 50 RU/mL			Matrix 100 RU/mL		
Spike d Conc.	Obs	Exp.	Obs/Ex p	Obs	Exp.	Obs/Ex p
Neat	47.5 7			102.3 9		
100 RU/m L	50.2 7	52.8 1	95.20%	100.9 8	102.1 5	98.90%
200 RU/m L	63.6 2	62.8 1	101.30 %	111.2 3	112.1 5	99.20%
300 RU/m L	71.2 2	72.8 1	97.80%	122.2 1	122.1 5	100.00 %

^{*}Representative data; results in individual laboratories may vary from these data.

TRACEABILITY

The reported values were determined with multiple runs on the BioCLIA® 1200 and BioCLIA® 6500 using specific reagents against an in-house standard. Results are reported in RU/mL, which is interpreted from relative light unit (RLU). Method comparison test showed good sensitivity and specificity.

PRECISION

A study based on guidance from CLSI document EP5-A2 was performed for determining the precision of BioCLIA DGP IgG kit. Human serum in the in-house reference panel (RP1, RP2, RP3, RP4) was tested with 10 replicates per sample for intra-assay precision evaluation, while with 4 replicates per sample for inter-assay precision. Each sample tested in individual runs, and 2 runs per day for 10 days, a total of 80 points. Data from this study are summarized in the following table.*

Intra-assay precision: CV< 10%

Intra-Assay	RP1	RP2	RP3	RP4
Mean(RU/ml)	10.77	20.19	99.79	359.96
cv	4.56%	4.62%	4.71%	4.28%

Inter-assay precision: CV < 15%

Inter-Assay	RP1	RP2	RP3	PR4
Mean(RU/ml)	10.41	19.39	99.25	357.09
cv	6.00%	5.71%	5.79%	4.30%

^{*}Representative data; results in individual laboratories may vary from these data.

LIMIT OF BLANK / DETECTION (LOB/LOD)

LOB/LOD was determined consistent with CLSI EP17-A guideline. LOB/LOD of the BioCLIA DGP IgG assay was lower than 1.0 RU/mL, which is below the analytical measuring range of the assay.

ASSAY REPORTABLE RANGE

The BioCLIA DGP IgG kit has a reportable linear range of 2 - 400 RU/mL. The linear range was determined by diluting a high positive anti-DGP IgG antibody serum sample with a negative sample to several concentrations which covers the entire assay linear range according to the scheme in CLSI EP6-A. The expected value was plotted against the observed value, and linear regression analysis was performed to get slope, intercept and coefficient of correlation (r) values. The results are summarized in the tables below: *

Slope	Intercept	r
0.98	+1.34	0.99

Assay linear range is 2-400 RU/mL. Results below the lower limit will be reported as < 2 RU/mL, while those are above the upper limit will be reported as > 400 RU/mL.

INTERFERENCE

Bilirubin, hemoglobin, triglycerides, rheumatoid factor (RF) and human anti-mouse antibody (HAMA) will not affect the BioCLIA DGP IgG assay performances when at the level indicated below.

Bilirubin ≤ 40 mg/dL;

Hemoglobin \leq 150 mg/dL;

^{*}Representative data; results in individual laboratories may vary from these data.

Triglycerides ≤ 1,000 mg/dL;

Rheumatoid factor (RF) ≤ 1,000 IU/mL;

Human anti-mouse antibody (HAMA) \leq 2,000 ng/mL.

METHOD COMPARISON

Method comparison was implemented by comparing BioCLIA DGP IgG assay to the predicated assay.

Clinical Sample		BioCLIA DGP IgG		
		-	+	Total
Predicated Method	-	48	2	50
	+	4	46	50
	Total	52	48	100

Sensitivity	92.0%
Specificity	96.0%
Total agreement	94.0%

Limitations

- The effectiveness of this kit is only confirmed for human serum/plasma, the applicability of the other kinds of samples is not verified.
- Reliable and reproducible results will be obtained when the assay procedure is carried out in accordance with the instructions and with adherence to good laboratory practice.
- Clinical diagnosis should not be made on the findings of a single test result, but should be integrate with all clinical and laboratory findings.

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HOB Biotech Group Co., Ltd

C6 Building, No. 218 Xinghu Road, Suzhoulndustrial Park,

Suzhou, Jiangsu, 215123, China

REGISTRANT/MANUFACTURE: HOB Biotech Group Co.. Ltd

ADDRESS/LOCATION:

C6 Building, No. 218 Xinghu Road, Suzhou Industrial Park, Suzhou, Jiangsu, 215123 China

CONTACT INFORMATION: TEL (+86)512-69561996

Fax (+86)512-62956652

WEBSITE: www.hob-biotech.com

CUSTOMER SERVICE: HOB Biotech Group Co., Ltd CUSTOMER SERVICE CONTACT: TEL (+86)4008601202



EUROPE REPRESENTATIVE: Emergo Europe **ADDRESS/LOCATION:**

Prinsessegracht 20, 2514 AP The Hague, The Netherlands

Technical Assistance

For technical assistance, contact your National Distributor.

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